

The Application of Biosurfactant Produced by *Azotobacter* sp. for Oil Recovery and Reducing the Hydrocarbon Loading in Bioremediation Process

Merry Sianipar, Edwan Kardena, and Syarif Hidayat

Abstract—Biosurfactant is potentially used to enhance oil recovery from oil sludge. The exopolysaccharide-formed biosurfactant is produced from *Azotobacter* sp. using 2 w/v% glucose sources. The oil sludge taken from company X and biosurfactant is agitated under the speed of 150 rpm and temperature of 20-25°C. The ratios of oil sludge and biosurfactant were varied by 1:1, 2:1 and 3:1. After 7 h agitation, the mixture was stood alone for 12 hours to obtain the 3 separated phases among oil (top), water (middle) and solid (bottom). The role of biosurfactant as surface tension reducer successfully separated among oil, water, solid from the oil sludge. The amounts of oil recovered from oil sludge are 55.95%, 51.76% and 25.57% for ratio 3:1, ratio 2:1, and ratio 1:1, respectively. Further, the slurry phase bioremediation method under 120 rpm of impeller velocity was applied to treat the bottom formation (slurry) from previous process. Both 5 w/v% Petrofilic bacteria (Unidentified Mix Culture) as a degrader agent and 5 w/v% fertilizer as nutrient were added in the first day of bioremediation process. Under controlled 10-40% solid concentration, 5.5-9.5 pH, 20-40°C (mesophilic condition), the total petroleum hydrocarbon (TPH) was successfully decreased until 6.83%, 3.48%, and 2.11% for slurry of ratio 1:1, ratio 2:1, and ratio 3:1 in 35 days.

Index Terms—*Azotobacter* sp., biosurfactant, oil recovery, slurry Phase bioremediation.

I. INTRODUCTION

Oil sludge caused from oil spill during transportation, separator tanks, cleaner tanks, and spray coolers is a pollutant that should be strictly concerned in petroleum industry. The oil sludge that cannot be returned to the earth by injection is only treated by mixed it with the soil from stockpile containing TPH.

Thus, the mixture soil is drying under the sunrise which are non-environmentally treatment leading to the damage of the ecosystem life. In this study, the issue of environment protection in the oil company X as one of the largest oil exploration companies in Indonesia is our focus. Based on

laboratory measurement, the oil sludge resulted by the company X contained TPH concentration over 20% which is indicating the presence of high loading pollutant which extremely impacts the environmental sustainability. Therefore, we offered an integrated approach for handling the oil sludge problem as environmental concerns: oil recovery process in the first step (pretreatment) followed by an ex-situ bioremediation process with slurry phase method in the second step (see Fig. 1). The oil recovery concept was offered due to the high concentration of TPH indicating the oil contained in oil sludge is still high. Therefore, it will be potentially economic. Furthermore, a pretreatment was applied to decrease the loading process of bacteria for bioremediation. This study used biosurfactant-produced *Azotobacter* sp. for pretreatment as enhancing oil recovery agent [1], [2]. Biosurfactant which basically produced from bacteria or microorganism [1], [3], [4] shows environmental compatibility and high activity at extreme temperatures, pH and salinity [5] is effective in decreasing the surface tension [6], critical middle concentration, interfacial tension in both aqueous solutions and hydrocarbon mixtures [7]

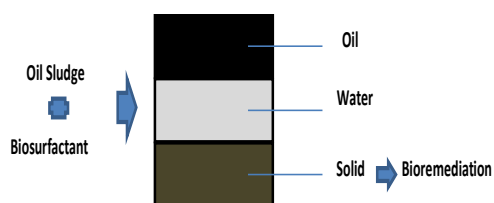


Fig. 1. Research concept.

II. MATERIALS AND METHODS

A. Oil Sludge Initiation

Oil sludge sample was taken from company X in Indonesia (see Fig. 2). Company X itself is an oil and gas company which produces heavy oil characteristic. Therefore, the oil contained in oil sludge was characterized as heavy oil which is indicated by the existence of three strong organic hydrocarbon including Pristine (nC_{17}), Phytane (nC_{18}) and Botryococcane (nC_{28}). Furthermore, the oil sludge characterization included: 25.02% of TPH (Total Petroleum Hydrocarbon), 7.38 of pH, 45-69% of moisture content, 867.8 kg/m³ for oil sludge density at 25°C, 450 centipoises (25°C) for viscosity, BTex analyse (EPA 8260B): 0 µg/L of benzene and ethylbenzene & xylene, 8.19 µg/L of toluena. We presumed that the existed toluene can be degraded in bioremediation process since *Pseudomonas* and *Achromobacter* which were reported as toluene degrader [7]

Manuscript received June 25, 2015; revised September 1, 2015. This work title is the Application of Biosurfactant produced by *Azotobacter* sp. for Oil Recovery and Reducing the Hydrocarbon Loading in Bioremediation Process.

Merry Sianipar is with Institut Teknologi Bandung (ITB) and Pasundan University, Bandung, Indonesia (e-mail: sianipar.merry.ms@gmail.com).

Edwan Kardena is with Institut Teknologi Bandung, Bandung, Indonesia. He is now with the Department of Environmental Engineering, Indonesia (e-mail: kardena@pusat.itb.ac.id).

Syarif Hidayat is with the Civil and Environmental Engineering Department, Hanyang University, Republic of Korea (e-mail: syarif_hidayat153@yahoo.com).

are included in our Petrofilic bacteria consortium.



Fig. 2. The oil sludge sample from company X.

B. Biosurfactant and Petrofilic Bacteria Preparation

The biosurfactant (see Fig. 3) was produced in our laboratory in accordance to our group previous reports [2], [8], [9]. Firstly, the *Azotobacter* sp. isolation was acclimated in Ashby Media using mannitol as the main carbon source [10]. Thus, Ashby Media as the selective media was agitated in 100 rpm speed for 72 h at 37°C which resulted in 10^6 cell/mL density bacteria. The Ashby Media (1 L DI water) contained mannitol (15 g), K_2HPO_4 (0.5 g), $MgSO_4 \cdot 7H_2O$ (0.2 g), $CaSO_4$ (0.1 g), NaCl (0.2 g), and $CaCO_3$ (5 g). Secondly, the acclimated *Azotobacter* sp. was proliferated and put into Basal Minimum Media to produce expolysaccharide-formed biosurfactant using 2% w/v glucose as carbon source. The basal minimum media (1 L DI water) contained K_2HPO_4 (1.5 g), KH_2PO_4 (0.5 g), $(NH_4)_2SO_4$ (0.4 g), $MgSO_4 \cdot 7H_2O$ (0.2 g), 10 mL trace element contained $Na_2EDTA \cdot 2H_2O$ (12 g), $FeSO_4 \cdot 7H_2O$ (2 g), $CaCl_2$ (1 g), $ZnSO_4 \cdot 7H_2O$ (0.4 g), Na_2SO_4 (10 g), $MnSO_4 \cdot 4H_2O$ (0.4 g), $CuSO_4 \cdot 5H_2O$ (0.1 g), and $Na_2MoO_4 \cdot 2H_2O$ (0.5 g) in 1 L DI water. The medium was sterilized in autoclave with temperature of 120°C and pressure of 15 psi for 20 min. Furthermore, the bacteria used in slurry phase bioremediation process was Petrofilic bacteria (Unidentified Mixed Culture) developed by the Environmental Biotechnology of Environmental Engineering of ITB [2], [9]. The Unidentified Mixed Culture bacteria was isolated consortium included *Pseudomonas stutzeri* BLO2, *Bacillus cereus* BL01, *Bacillus* sp. BL04, and *Achromobacter* sp BL03.



Fig. 3. Biosurfactant produced by *Azotobacter* sp.

C. Oil Recovery and Bioremediation Preparation

The ratio variations between biosurfactant and oil sludge were 3:1, 2:1 and 1:1. The mixture was agitated in the agitator reactor under 150 rpm speed, 7 hours and at room temperature (20-25°C) [1]. Moreover, for bioremediation process, slurry phase bioreactor (see Fig. 4) was used and set in the range of 100-200 rpm speed as optimum mixture speed [11]. The dose ratio of additional Petrofilic bacteria was 5w/v% [12].

D. Materials Characterization

As In this study, the total petroleum hydrocarbon (TPH)

concentration was the main parameter used to observe the amount of recovered oil and the TPH decrease during both oil recovery as well as bioremediation process. TPH was analyzed by extraction gravimetric (modification of EPA methods 164 Rev A). 15 ml vial was heated in oven at 105°C for 1 h, then, put into desiccator for cooling. The cooled vial was weighted called (a). Hexane was put into the erlenmeyer as much as two times volume of the sample to be examined. The weight of the sample was called (c). The erlenmeyer was stirred at 150 rpm for 2 h in orbital shaker. The erlenmeyer was stood alone for 1 h until formed 2 separate phases. The top phase which is the solution of N-Hexane and extracted oil put in 15 ml vial. The filled vial was heated in a water bath at 70°C to evaporate the N-hexane. Further, that vial was cooled back at desiccator and weighted (b). The equation to calculate TPH based on extraction gravimetric method is shown on Equation 1. Moreover, to describe the oil behavior of sample during process, the oil fingerprint of sample was measured (GC-5890). Furthermore, the characteristics of recovered oil were investigated using oil fingerprint (GC-5890), and density & API number (hydrometer method-ASTM D4052). In addition, other parameters such as solid concentration, pH and Temperature were checked to observe the optimum condition during treatment. Solid concentration was expected in the range of 10-40% to enhance the biodegradation process [11]. Meanwhile pH and temperature was determined using pH/Temperature meter (model 410A, 110 VAC).

$$TPH(\%) = [(b - a) / c] \times 100 \quad (1)$$

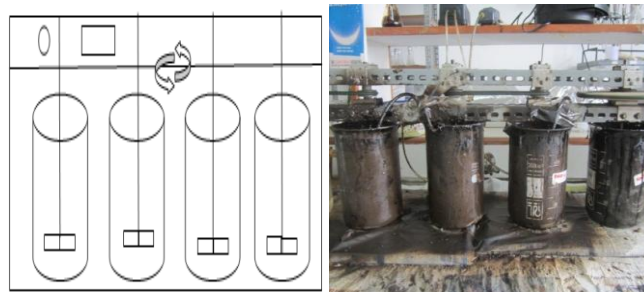


Fig. 4. Slurry phase bioreactor.

III. RESULTS AND ANALYSIS

A. Recovered Oil Characterization

The separated phase formed after mixing biosurfactant and oil sludge were shown on Fig. 5. The recovered oil was found at 140 ml for ratio 3:1, 130 ml for ratio 2:1 and 64 ml for ratio 1:1. The highest oil recovery was achieved by the ratio 3:1 while the lowest was achieved by ratio 1:1. It was caused by the stronger separation given by the increase of biosurfactant amount. Moreover, since the results of ratio 2:1 revealed insignificant difference comparing with the results of ratio 3:1, we presumed that the ratio 2:1 was the optimum ratio in oil recovery process. Furthermore, the characteristic of oil showed that the recovered oil characteristic was not affected by the biosurfactant. The oil fingerprint and API & density number data of the oil recovered showed the heavy oil characteristic [13] as the original characteristic oil of Company X (see Fig. 6, Table I and Fig. 7).



Fig. 5. The separated phase formed after mixing biosurfactant and oil sludge at the ratio 1:1 (Left) and the ratio 2:1 (Right) with following oil on the top, water in the middle and solid in the bottom.

TABLE I: THE QUALITY OF RECOVERED OIL

Parameters	Unit	Ratio 1:1	Ratio 2:1
Density	g/cm ³	0.93225	0.93539
Actual Temperature	°C	40	40
Rho API 60°F-C	g/cm ³	0.9482	0.9512
API 60 °F-C	-	17.59	17.1
SG 60 °F-C	-	0.9491	0.9521
Pour Point	°F	40	35

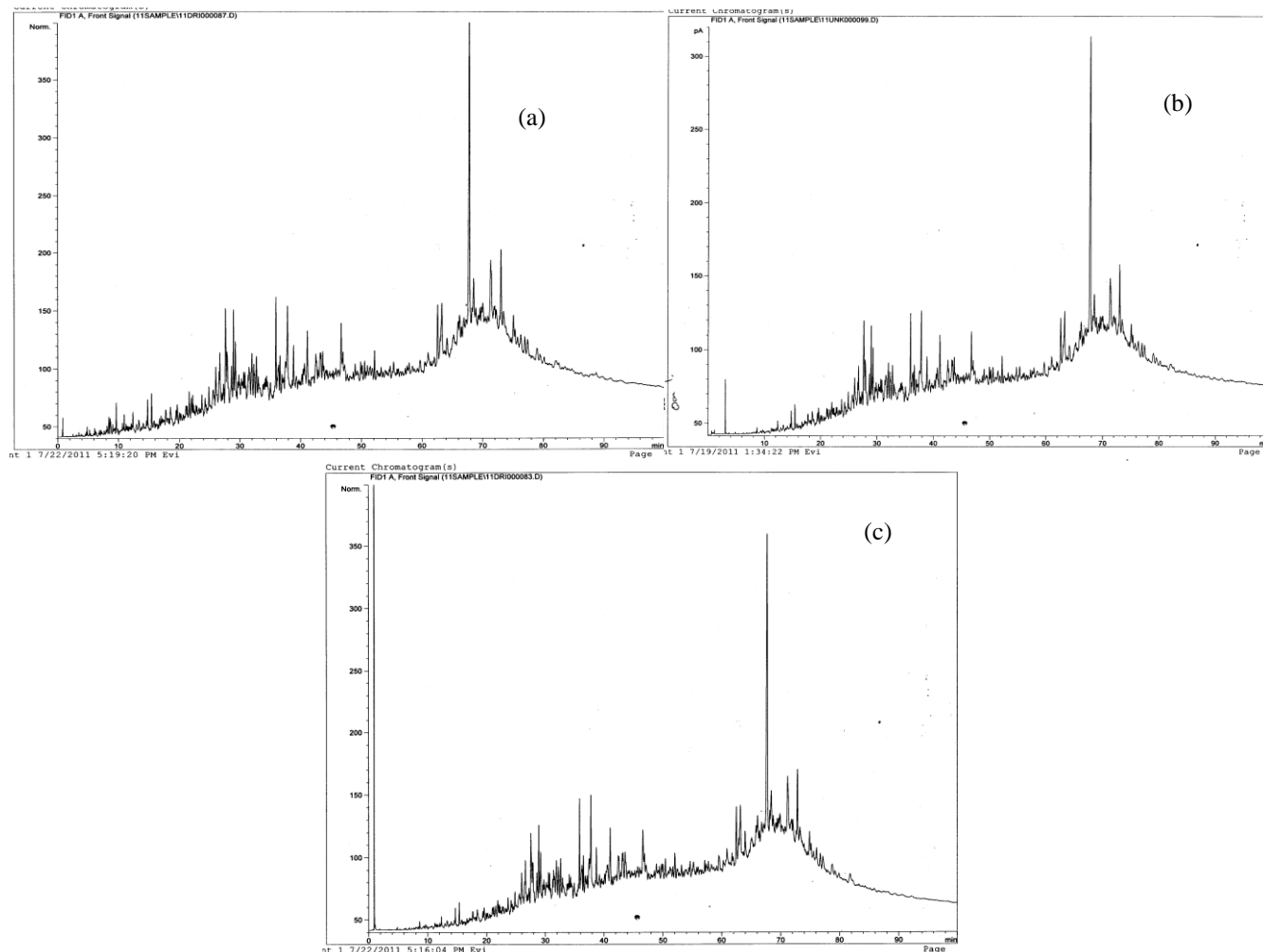


Fig. 6. The oil fingerprint of recovered oil from ratio 1:1 (a), ratio 2:1 (b), and Company X (c).

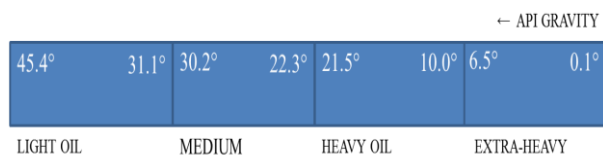


Fig. 7. API standardization referred to American Petroleum Institute [13].

A. Solid Phase (Slurry) Characterization and Slurry Phase Bioremediation Treatment

The bottom formations (solid phase) were characterized and found at 18.6% TPH for ratio 1:1, 12.1% TPH for ratio 2:1, and 11% TPH for ratio 3:1. It was consistent with the amount of recovered oil data showing the ratio 3:1 and ratio 2:1 had higher oil recovery than ratio 1:1. Moreover, the pH of slurry, around 5.6-5.9 pH for all ratios, was decreased from the initial condition, i.e., pH 7.38, due to the work of bacteria in [11] during the oil recovery process. Furthermore, the high

moisture (%) of solid phase result, more than 60% moisture, for all ratios, indicated that the liquid concentration was highly contained on solid phase called slurry. Therefore, the slurry phase bioremediation was chosen to treat solid phase. Furthermore, during bioremediation, the total petroleum hydrocarbon (TPH) was decreased for pretreated slurry (see Fig. 8). Moreover, the pristine, phytane and botryococcane identified as strong organic hydrocarbon, was successfully treated in bioremediation process described by oil fingerprint results (see Fig. 9). The decrease of TPH was caused by two factors: bacteria in bioremediation process and the biosurfactant role. The role of Unidentified Mixed Culture bacteria or Petrofilic bacteria strongly biodegraded the hydrocarbon oil in slurry [12]. Moreover, the percentages of organic compound contained in solid (see Fig. 10) showed that bacteria preferred eat the light fraction first than the heavy fraction before. In addition, the smallest portion of light

fraction especially at initial condition was caused by the light hydrocarbon vaporize [1]. The form of dried slurry after bioremediation process and without bioremediation process (control) could be seen on Fig. 11.

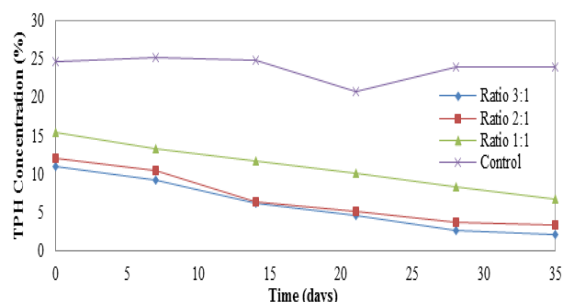


Fig. 8. TPH decrease during bioremediation process and oil fingerprint of slurry after bioremediation process field.

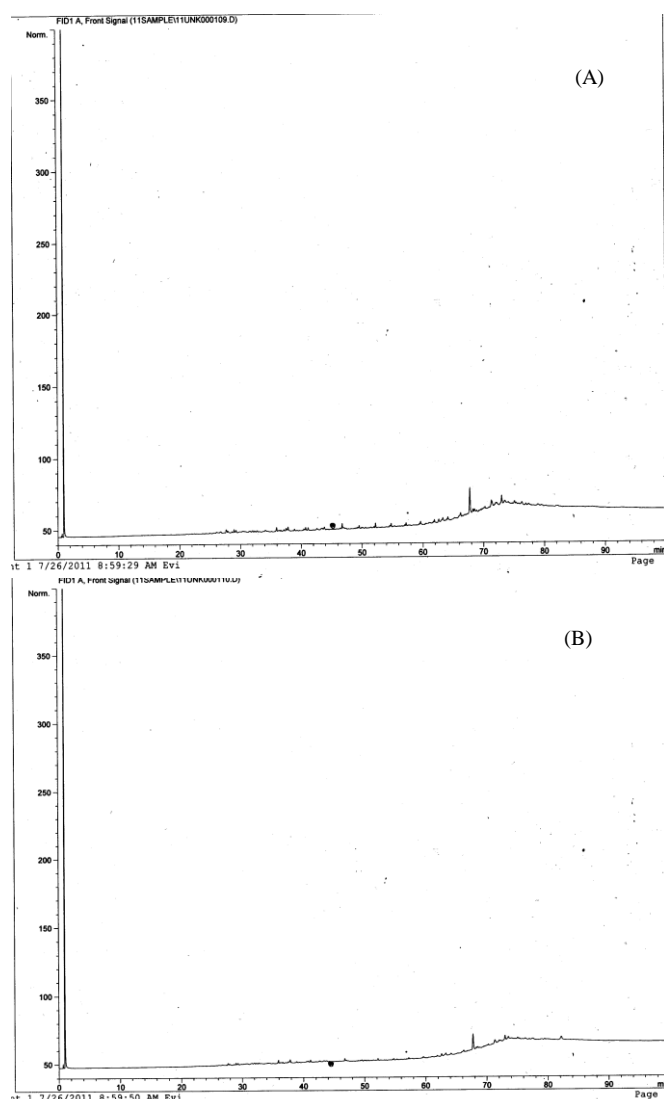


Fig. 9. The oilfingerprint of slurry after bioremediation process for Ratio 1:1 (A) and Ratio 2:1 (B).

Furthermore, the use of biosurfactant in pretreatment was presumed to give influence to the oil degradation. As emulsifier agent, biosurfactant gave arise soluble oil that led to the easiness for bacteria in degrading the hydrocarbon (bioavailability) [14]. Moreover, it was showed that the more used biosurfactant, the more increase the bioavailability of hydrophobic compound [14], [15]. Controlling the solid concentration, temperature, and pH in slurry phase bioreactor

(see Fig. 12) also contributed for successful biodegradation [11], [16].

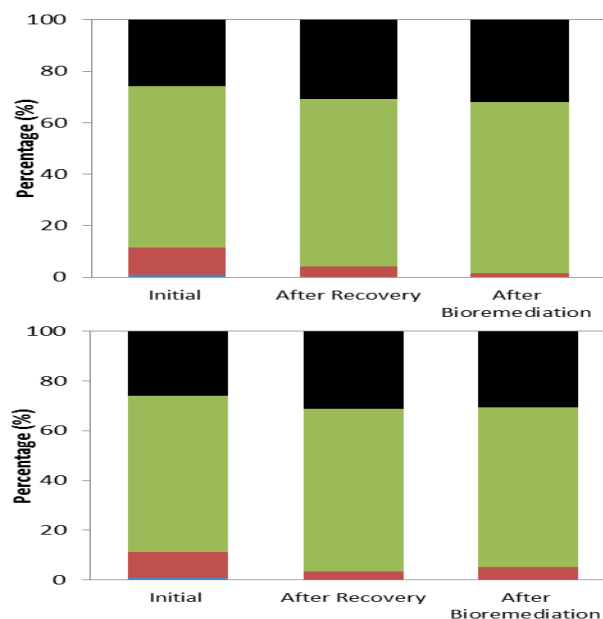


Fig. 10. The percentage of organic compound composition in solid (C6-C9, blue; C10-C24, red; C15-C28, green; C29-C36, black) from Ratio 1:1 (Top) and Ratio 2:1 (Bottom).



Fig. 11. The digital photos of the dried slurry after bioremediation process (left to right: Control, Ratio 1:1, Ratio 2:1, Ratio 3:1).

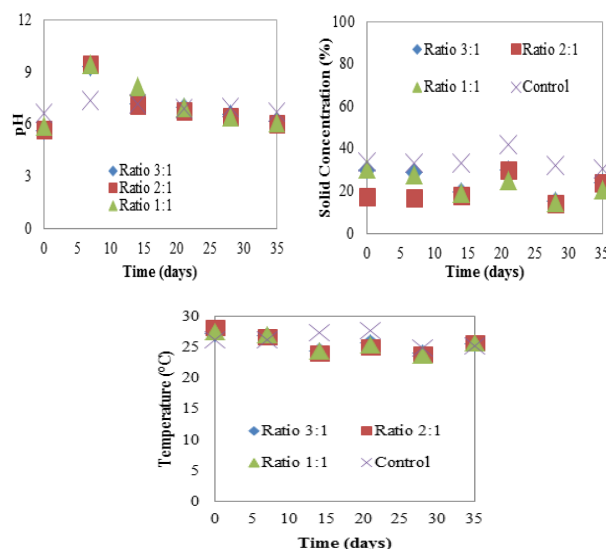


Fig. 12. Solid concentration, temperature, and pH of slurry during bioremediation process.

IV. CONCLUSION

Biosurfactant produced by *Azotobacter* sp. was found to be an oil emulsifier to separate oil, water, and solid from oil sludge. Moreover, biosurfactant is feasible in recovering oil from oil sludge without bothering the quality of recovered oil. Ratio 2:1 was the optimum dose ratio in the oil recovery process. Furthermore, the decreased TPH of recovered solid from the pretreatment process was further treated in the slurry phase bioremediation process using *Petrofilic* bacteria. The

order of TPH decrease was ratio 3:1 > ratio 2:1 > ratio 1:1. The Petrophilic bacteria plays role in decreasing the TPH of the slurry. Considering the feasibility and performances, we recommended the ratio 2:1 as the optimum ratio for obtaining the optimum oil recovery and bioremediation performance.

ACKNOWLEDGMENT

Authors give gratitude to PT Chevron Pacific Indonesia, Riau, Indonesia, for allowing Authors to analyze our samples for some parameters including API, TPH, pour point, oil fingerprint and BTex parameters in the Petroleum Engineering Laboratory.

REFERENCES

- [1] C. Zheng, L. Yu, and L. Huang, "Microbial enhanced treatment of oil sludge from oil production plant by rhodococcus ruber Z25," *Journal of International Canadian Petroleum Technology*, vol. 51, pp. 290-294 2012.
- [2] Q. Helmy, E. Kardena, Z. Nurachman, and Wisjnuaprpto, "Application of biosurfactant produced by azotobacter vinelandii AV01 for enhanced oil recovery and biodegradation of oil sludge," *International Journal of Civil & Environmental Engineering (IJCEE)*, vol. 10, pp. 7-1, 2010.
- [3] I. M. Banat, N. Samarah, and M. Murad, "Biosurfactant production and use in oil tank clean-up," *World J. Microbial Biotechnol.*, vol. 7, pp. 80-88, 1991.
- [4] E. J. Gudina, L. R. Rodrigues, J. A. Teixeira, J. F. Pereira, and J. A. Coutinho, "Biosurfactant producing microorganisms and its application to enhance oil recovery at lab scale," presented at the SPE (Society of Petroleum Engineers) EOR Conference at Oil and Gas West Asia, Muscat, Oman, 2012.
- [5] C. C. Lai, Y. C. Huang, Y. H. Wei, and J. S. Chang, "Biosurfactant-enhanced removal of total petroleum hydrocarbons from contaminated soil," *Journal of Hazard Mater.*, vol. 167, no. 1-3, pp. 609-14, 2009.
- [6] R. D. Rufino, J. M. de Luna, G. M. T. Takaki, and L. A. Sarubbo, "Characterization and properties of the biosurfactant produced by Candida lipolytica UCP 0988," *Electronic Journal of Biotechnology*, vol. 17, pp. 24-38, 2014.
- [7] D. Claus, "The decomposition of toluene by soil bacteria," *Journal of Gen Microbiol.*, vol. 36, pp. 107-22, 1964.
- [8] Q. Helmy, E. Kardena, N. Funamizu, and Wisjnuaprpto, "Optimization of cultural conditions for biosurfactant production from azotobacter vinelandii," presented at the International Conference on Sustainable Environmental Technology and Sanitation for Tropical Region in Surabaya, Indonesia, 2008.
- [9] Q. Helmy, Wisjnuaprpto, and E. Kardena, "The effect of mixing velocity gradient in bio-emulsifier production by azotobacter sp. Used for biodegradation of petroleum hydrocarbon," presented at the 1st Annual Congress of Oil Field Chemicals, Beijing China, 2010.
- [10] Schlegel and G. Hans, *General Microbiology*, p. 206, England: Cambridge University, 1986.

- [11] R. L. Irvine and S. K. Sikdar, *Bioremediation Technologies Volumes III*, pp. 464-465, USA: Technomic Publishing Company, 1998.
- [12] Q. Helmy, R. Laksmoni, and E. Kardena, "Bioremediation of aged petroleum oil contaminated soil: From laboratory scale to full scale application," presented at the 2nd Humboldt Kolleg in conjunction with International Conference on Natural Sciences (HK-Icons), 2014.
- [13] Center for energy. [Online]. Available: www.centerforenergy.com
- [14] E. Z. Ron and E. Rosenberg, "Biosurfactant and bioremediation," vol. 13, pp. 249-252, 2002.
- [15] R. Boopathy, "Factors limiting bioremediation technologies," *Bioresource Technology*, vol. 74, pp. 63-67, 2000.
- [16] G. V. Weert, D. V. D. Werff, and J. J. Derksen, "Transfer of O₂ from air to mineral slurries in a Rushton turbine agitated tank," *Minerals Engineering*, vol. 10, pp. 1109-1124, 1995.



Merry Sianipar comes from Pematangsiantar, Indonesia. She received the B.S. in Environmental Engineering Department from Institut Teknologi Bandung (ITB), Indonesia in 2012 and the M. S. degree from Kyungnam University, Republic of Korea in 2015. She had been engaged on research of nanocomposite membrane technology for water treatment application.. She is now affiliated with Institut Teknologi Bandung (ITB) as a research assistant in Downstream Process Laboratory and Pasundan University as a lecturer, Bandung, Indonesia. Her research interests include drinking water treatment, water supply, nanocomposite membrane, and environmental biotechnology.



Edwan Kardena comes from Bandung, Indonesia. He received the B.Eng. in Environmental Engineering Department from Institut Teknologi Bandung (ITB), Indonesia in 1988 and the Ph.D degree in the field of environmental microbiology/biotechnology from University of Wales College of Cardiff in United Kingdom in 1995. He is currently working as a lecture as well as a researcher at Environmental Engineering Dept. of Institut Teknologi Bandung, Indonesia. He has been engaged on research and development of microbiology/biotechnology especially in biological treatment processes, bioremediation and bio-energy. His research interests include water and waste water treatment, microbiology and environmental biotechnology.

Dr Kardena has involved in the WJEMP-LES Kota Bogor as expert for wastewater/sanitation. He is the author of many papers over 15 papers.



Syarif Hidayat comes from Bandung, Indonesia. He received the B.S. in environmental engineering from Institut Teknologi Bandung (ITB), Indonesia in 2009 and the M.S. degree at the same university in 2012. He is currently pursuing the Ph.D. degree at Civil and Environmental Engineering Department of Hanyang University, Republic of Korea. He has been engaged research and development of domestic and industrial wastewater treatment technologies especially in biological treatment processes and bio-energy. His research interests include microbial fuel cell, microbial electrolysis cell and microbial reverse-electrodialysis electrolysis cell.